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expression, and purification procedure of recombinant HBsAg elicits immunogenic response in mice indicating its therapeutic potential.

Recombinant streptokinase (rSK) was synthesized by removing the bacterial signal sequence and fusing the structural gene encoding streptokinase to a constitutive promoter of glyceraldehyde-3-phosphate dehydrogenase (GAP) gene in Pichia pastoris. *P. pastoris* strains that expressed rSK produced the protein in an intracellular form. A 47-kDa protein was produced that reacted with polyclonal antibodies raised against native streptokinase. SK produced by constitutive expression was found to be functionally active by plasminogen activation assay and clot lysis assay. Continuous fermentation strategy with YPD medium was employed successfully with no toxicity to host. Recombinant streptokinase expressed was partially purified by DEAE-Sepharose/Phenyl Sepharose chromatography with no endotoxins. The present cloning, continuous expression, and purification procedure of recombinant streptokinase from *P. pastoris* enables preparation of large quantity of recombinant streptokinase for structure, function studies and evaluation of its clinical potential in thrombolytic therapy.