

# **CHAPTER-III**

**18s rRNA gene polymorphisms of *P. homarus* populations from different geographic regions**

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## INTRODUCTION

The spiny lobster *P. homarus* has long been of interest to evolutionary biologists because of its high level of species diversity, its wide geographic distribution, and the importance of many species to commercial fisheries. Among genera in the family Palinuridae, *Panulirus* has been the most successful in terms of species diversity; 19 species have been described to date, three of which are divided into seven recognized subspecies (Holthuis 1991; George 1997; Sarver *et al.*, 1998). The key to this successful radiation is thought to be the invasion by species of *Panulirus* of shallow-water, tropical environments, which permits the occupation of varied habitats not accessible to congeners (George and Main 1967; George 1997). Because of morphological, ecological, and behavioural diversity, both within and among species of *Panulirus*, the phylogenetic history of this group has not been well understood.

The 16S rRNA sequences (16S) have been one of the most common segments of the mitochondrial genomes used in phylogenetic analysis. For crustaceans they have been used in the analysis of penaeid prawns (Palumbi and Benzie, 1991; Bouchon *et al.*, 1994), freshwater crayfishes (Grandjean *et al.*, 1998; Crandall *et al.*, 1999), marine lobsters (Harding *et al.*, 1997; Ovendon *et al.*, 1997; Sarver *et al.*, 1998; Machordom and Macpherson, 2004), fairy shrimps (Daniels *et al.*, 2004), Caribbean sponge-dwelling snapping shrimps (Morrison *et al.*, 2004), and freshwater prawns (Murphy and Austin, 2002,

2003, 2004, 2005; Liu *et al.*, 2007). A few recent studies indicated that the 16S produced poor resolution in reconstructed phylogenies for insects, perhaps due to its fast evolutionary rate (Misof *et al.*, 2001; Hasegawa and Kasuya, 2006). The poor resolution was also found for phylogenies of freshwater prawns. Murphy and Austin (2005) made the phylogenetic analysis of 30 species of *Macrobrachium*, but it resulted in a poor resolution; their phylogenetic tree had a very high degree of polytomy represented by a single multifurcating node with the clades of 28 taxa, 93% of the total taxa under the study. Liu *et al.*, (2007) increased the number of taxa to 38 but also obtained a poor resolution; their phylogenetic tree also had a high degree of polytomy represented by a multifurcating node with the clades of 37 taxa, 97% of the total taxa under the study.

Recently, nuclear 18S rRNA sequences (18S) have been used in phylogenetic studies of crustaceans (Crandall *et al.*, 2000; Jarman *et al.*, 2000). The 18S gene of nuclear DNA was found to have an evolutionary rate similar to or slower than that of the 16S gene (Fetzner and Crandall, 2001). It was able to clarify the relationships among distantly related taxa for water striders (Muraji and Tachikawa, 2000) and Odonata (Hasegawa and Kasuya, 2006), and also among closely related taxa for insects in Ichneumonoidea (Belshaw *et al.*, 1998) and the hominoids (Gonzalez *et al.*, 1990).

This study is intended to determine whether the 18S is a better tool than the 16S in phylogenetic analysis of spiny lobsters collected from five different geographical regions, by comparing their characters and evaluating topological resolution and phylogenetic signals of their reconstructed phylogenetic trees.

## Materials and Methods

### DNA extraction and Primer design

Total genomic DNA was extracted using a modified CTAB protocol as described in Chapter I. The 18S lobster primer sequences and their GC contents were given in table: 6. 1  $\mu$ M concentration of primer was used in PCR as well as sequencing. The protocols were followed according to the manufacturer's directions.

**Table: 6. 18S rRNA Sequence primer.**

Primer	Sequence	Tm	%GC	Length(bp)
Forward 5'	CGCACGAGAATGAGCAATAA	59	45	20
Reverse 3'	GTACAAAGGGCAGGGACGTA	59	55	20

The portion of the 18S gene was amplified by PCR (polymerase chain reaction) with above primer. Double-stranded PCR products were obtained in a total reaction volume of 50  $\mu$ L, containing 5  $\mu$ L of 103 ExTaq buffer (Mg<sup>2+</sup> added; TaKaRa), 5  $\mu$ L of dNTP mixture (2.5 mM of each), 1  $\mu$ M of each primer, 1 unit of Taq polymerase (TaKaRa Ex Taq<sup>TM</sup>; BIOGENO, INDIA), and 5  $\mu$ L of DNA extract (100 ng). For the 18S sequences, the target segment was amplified by PCR in the following temperature regime: the initial denaturizing step at 94<sup>o</sup>C for 5 min, followed by 35 cycles of denaturizing at 94<sup>o</sup>C for 1 min, an annealing temperature of 48–55<sup>o</sup>C for 1 min, an extension temperature of 72<sup>o</sup>C for 1 min, and then an additional extension of 72<sup>o</sup>C for 5 min.

For the 18S sequences, PCR products were purified using Geneaid Genomic DNA Purification Kit (ORCUTIS SCIENTIFIC). All sequencing reactions followed the Perkin Elmer protocol, using a BigDye Terminator v3.1 Cycle Sequencing Kit with a DNA Analyzer (APPLIED BIOSYSTEMS). The sequencing was performed in both directions. The forward and reverse sequence chromatograms were checked and edited manually using the programs of SeqMan 5.01 (DNASTAR) and BioEdit version 7.0.4.1 (Hall, 1999). Multiple alignments were performed using Clustal W (Thompson *et al.*, 1994) with default parameter values (gap open penalty 10, gap extension penalty 0.1 in pairwise and 0.05 in multiple alignments) and all alignments were manually checked. All sequences of the 18S extracted in this study were deposited at the GenBank database (Table 7).

### **Sequence Character Analysis**

Characters of nucleotide sequences were examined with the exploratory data analysis, using MEGA version 3.1 (Kumar *et al.*, 2004), DAMBE version 50.7 (Xia and Xie, 2001) and DnaSP 4.00 (Rozas *et al.*, 2003). The genetic divergent distances among taxa were calculated using the best appropriated DNA substitution model obtained from Modeltest version 3.7 (Posada and Crandall, 1998). The distribution of pair-wise sequence divergences among the *P. homarus* sequences was established using SigmaPlot version 8.0.

True evolutionary relationships would be obscured in DNA sequence data sets if sites have become saturated by multiple substitutions (Swofford *et al.*, 1996; Jesus *et al.*, 2007). In order to assess the level of saturation in 18S rRNA gene, pair-wise sequences comparisons were performed by plotting the number of nucleotide substitutions per site for transitions and transversions, respectively, against the genetic divergent distances calculated from the best

appropriated DNA substitution model, using PAUP\* 4.0b10 (Swofford, 2002) and SigmaPlot version 8.0.

## RESULTS

Characters of the extracted 18S gene sequence of five different sites are shown in Plate: 1. The data has more or less same alignment length with 1798 nucleotide bases and were 0.01% in both variable and parsimoniously informative sites. Also, there was a significant difference in the percentage composition of nucleotide contents (adenine, thymine, cytosine, and guanine) between the collection sites. A significant difference was also found in relative frequencies of nucleotide substitutions (adenine-guanine, cytosine-thymine, adenine-cytosine, adenine-thymine, cytosine-guanine, and guanine-thymine).

The 18s rRNA gene was amplified and sequenced by using primer Forward 5'CGCACGAGAATGAGCAATAA and reverse 3'GTACAAAGGGCAGGGACGTA. The sequenced 18S rRNA genes were deposited on Genbank (Table: 7).

**Table: 7. Genbank Accession Number and Length of Gene Sequence.**

Species	Location	Gene	Genbank Accession No.	Gene sequence Length.
<i>P.homarus</i>	Vizhinjam	18S	GQ411528	1799 bp.
<i>P.homarus</i>	Cochin	18S	GQ411529	1798 bp.
<i>P.homarus</i>	Chinnamuttom	18S	GQ411530	1797 bp.
<i>P.homarus</i>	Tuticorin	18S	GQ411531	1800 bp.



Plate: 1

a) 18s rRNA sequence of *Panulirus homarus* from Vizhinjam

ATTAAGGCGAAACCGCGAATGGCTCATTAATCAGCTATGATTTCATTGGATCTGTAAACCCACTTACTTGGATAACTGTGGTAATT  
CTAGAGCTAATACATGCATCACGTCTCTGACCGCAAGGGAAGAGCGCTTTTATTAGTTCAAACCCAGTCGGGCCTCGGTCCGTCAC  
CCACCTGTGGTGAATCTGAATAACTTCTCGCTGTAGCCAGTGTCTACGCACCGGCCCGAGTCTTTCAAGTGTCTGCCTTATCAGC  
TTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTAACGGGGAATCAGGGTTCGATTCCGGAGAGGGAGCCTGAGAAACG  
GCTACCACATCTAAGGGAGGCAGCAGGCACGCAAATTACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAACGATGCGAGAC  
TCATCCGAGGCCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGAGGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCG  
CGGTAATTCAGCTCCAAATAGCGTATATTAAGTTGTTGCGGTTAATAAGCTCGTAGTTGGATTTCAGTTCGGACTGACGGTTCA  
CCGCCCGGTGTCTACTGTACGCTCCGAACAGCCGACCGCCGCTCGCACGGGGTGTCTTTCATCGAGTGTCCCGAGTGGCCGGC  
ACGGTTATTTGAAAAAATTAGAGTGTCTCAGAGCAGGCTCCTTGAATGGCCTGAATGTCTATGCATGGAATAATGGAATAGGACCT  
CGGTTCTATTTTGTGGTTTTCGCGAACCAGGTAATGACTAATAGGAATTGGCGGGTTCATTTCGATTTCGACGCTAGAGGTGAA  
ATTCTTGGACCGTCGCAAGCAACTACTGCGAAAGCAATTTGCCAAGGATGTTTCATTAATCAAGAACGAAAGTTAGAGGTTGAA  
GGCGATCAGATAACCGCCCTAGTTCTAACCATAAACGATGCTGACTAGCGATCCGCCGCGCTTATTCCCATGACCCGGCGGGCAGCT  
TCCGGGAAACCAAAGTCTTTGAGTTCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAAGGGCACCACCAGG  
AGTGGAGCCTGCGGCTTAATTTGACTCAACTCGGGGAACCTCACCAGGCCCGGACACCGGAAGGATTGACAGATTGAGAGCTCTTT  
CTCGATTTCGGTGGTGGTGGTGCATGGCCGTTCTTAGTTGGTGGAGCGATTTGTCTGTTAATTCGGATAACGAACGAGACTCTAG  
CCTATTAACCTAGTCGACGGATCTCCAGCATGCTCCGTGGTGTCCGTTGCAACGCTCTTCTAGAGGGATAAGCGGCAATTTCTAGCC  
GCACGAGAAATGAGCAATAACAGGTCTGTGATGCCCTTAGATGTTCTGGGCCGCACGCGCCTACACTGAAGGGATCAACGTGTTCT  
CCCCCTCCGAGAGGAGCGGGTAACCGATCAAACCCCTTCATGATAGGGATTGGGGCTTGCAATTTGTTCCCATGAACGAGGAATT  
CCCAGTAAGCGCAAGTCAATCAGCTTGCCTTGATTTCTCCCTGCCCTTTGTACACACCGCCCGTCTACTACCGATTGAATGATT  
TAGTGAGGCCTTCGGACTGGCGCTCTGGATGTTCTACCCCATCCCTTCTATTCTCGCAAGGGTCTGGAGGTTGGGGTCTCGCC  
TCGAGCTGACGGAAGATGTCCAAACTTGATCATCTAGAGGAAGTAAAGTCGTACCAAGGTTTCCGTAGGTGAACCTGC

b) 18s rRNA sequence of *Panulirus homarus* from Cochin

ATTAAGGCGAAACCGCGAATGGCTCATTAATCAGCTATGATTTCATTGGATCTGTAAACCCACTTACTTGGATAACTGTGGTAATT  
CTAGAGCTAATACATGCATCACGTCTCTGACCGCAAGGGAAGAGCGCTTTTATTAGTTCAAACCCAGTCGGGCCTCGGTCCGTCAC  
CCACCTGTGGTGAATCTGAATAACTTCTCGCTGAGCGCAGCGTTTACGCACCGGCCCGAGTCTTTCAAGTGTCTGCCTTATCAGC  
TTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTAACGGGGAATCAGGGTTCGATTCCGGAGAGGGAGCCTGAGAAACG  
GCTACCACATCTAAGGAAGGCAGCAGGCACGCAAATTACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAACGATGCGAGAC  
TCATCCGAGGCCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGAGGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCG  
CGGTAATTCAGCTCCAAATAGCGTATATTAAGTTGTTGCGGTTAAAAAGCTCGTAGTTGGATTTCAGTTCGGACTGACGGTTCA  
CCGCCCGGTGTCTACTGTACGCTCCGAACAGCCGACTTCCGGCTCGCACGGGGTGTCTTTCATCGAGTGTCCCGAGTGGCCGGC  
ACGTTTACTTTGAAAAAATTAGAGTGTCTCAGAGCAGGCTCCTTGAATGGCCTGAATGTCTATGCATGGAATAATGGAATAGGACCT  
CGGTTCTATTTTGTGGTTTTCGCGAACCAGGTAATGACTAATAGGAACAGGCGGGGCATTTCGATTTCGACGCTAGAGGTGAA  
ATTCTTGGACCGTCGCAAGACAACCTACTGCGAAAGCAATTTGCCAAGGATGTTTTTCATTAATCAAGAACGAAAGTTAGAGGTTGAA  
GGCGATCAGATAACCGCCCTAGTTCTAACCATAAACGATGCTGACTAGCGATCCGCCGCGCTTATTCCCATGACCCGGCGGGCAGCT  
TCCGGGAAACCAAAGTCTTTGAGTTCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAAGGGCACCACCAGG  
AGTGGAGCCTGCGGCTTAATTTGACTCAACCGGGGAACCTCACCAGGCCCGGACACCGGAAGGATTGACAGATTGAGAGCTCTTTC  
TCGATTTCGGTGGTGGTGGTGCATGGCCGTTCTTAGTTGGTGGAGCGATTTGTCTGTTAATTCGGATAACGAACGAGACTCTAGC  
CTATTAACCTAGTCGACGGATCTCCAGCATGCTCCGTGGTGTCCGTTGCAACGCTCTTCTAGAGGGATAAGCGGCAATTTCTAGCCG  
CACGAGAAATGAGCAATAACAGGTCTGTGATGCCCTTAGATGTTCTGGGCCGCACGCGCCTACACTGAAGGGATCAACGTGTTCTC  
CCCCCTCCGAGAGGAGCGGGTAACCGATCAAACCCCTTCATGATAGGGATTGGGGCTTGCAATTTGTTCCCATGAACGAGGAATTC  
CCAGTAAGCGCAAGTCAATCAGCTTGCCTTGATTACGTCCTGCCCCTTTGTACACACCGCCCGTCTACTACCGATTGAATGATTT  
AGTGAGGCCTTCGGACTGGCGCTCTGGATGTTCTACCCCATCCCTTCTATTCTCGCAAGGGTCTGGAGGTTGGGGTCTCGCCT  
CGAGCTGACGGAAGATGTCCAAACTTGATCATTTAGAGGAAGTAAAGTCGTACCAAGGTTTCCGTAGGTGAACCTGC

c) 18s rRNA sequence of *Panulirus homarus* from Muttom

ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCAATGGATCTGTAAACCCACTTACTTGGATAACTGTGGTAATT  
CTAGAGCTAATACATGCATCACGTCTCTGACCGCAAGGGAAGAGCGCTTTTATTAGTTCAAAACCCAGTCGGGCCTCGGTCCGTAC  
CCACCTGTGGTGAATCTGAATAACTTCTCGCTGAGCGCAGGGTCTACGCACCGGCCCGAGTCTTTCAGTGTCTGCCTTATCAGC  
TTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTAACGGGGAATCAGGGTTCGATTCCGGAGAGGGAGCCTGAGAAACG  
GCTACCACATCTAAGGAAGGCAGCAGGCACGCAAATTAACCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAACGATGCGAGAC  
TCATCCGAGGCCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGAGGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCG  
CGGTAATCCAGCTCCAATAGCGTATATAAAGTTGTTGCGGTTAAAAAGCTCGTAGTTGGATTTTCAGTCCCGACTGACGGTTCA  
CCGCCCGGTGTCTACTGTACGCTCCGAACAGCCGCACCTCCGGCTCGCACGGGGTGTCTTTCATCGAGTGTCCCGAGTGGCCGGC  
TCGTTTACTTTGAAAAAATTAGAGTGTCTCAGAGCAGGCTCCTTGAATGGCCTGAATGTCTATGCATGGAATAATGGAATAGGACCT  
CGGTTCTATTTTGTGGTTTACGGAACCCGAGGTAATGACTAATAGGAACAGGCGGGGCATTTCGTATTGCGACGCTAGAGGTGAA  
ATTCTTGGACCGTTCGCAAGACGAACTACTGCGAAAGCAATTTGCCAAGGATGTTTTCATTAATCAAGAACGAAAGTTAGAGGTTCGAA  
GGCGATCAGATACCGCCCTAGTTCTAACCATAAACGATGCTGACTAGCGATCCGCCGGCGTATTCCCATGACCCGGCGGGCAGCT  
CCGGAAACCAAGTCTTTGAGTTCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAAGGGCACCACAGG  
AGTGGAGCCTGCGGCTTAAATTTGACTCAACTCGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAAGGGCACCACAGG  
CTCGATTCCGGTGGGTGGTGGTGCATGGCCGTTCTTAGTTGGTGGAGCGATTGTCTGGTTAATTCGGATAACGAACGAGACTCTAG  
CCTATTAAGTGTGACGGATCTCCAGCATGCTCCGTGGTGTCCGTTGCAACGTCTTCTTAGAGGGATAAGCGGCAATCTAGCC  
GCACGAGAAATGAGCAATAACAGGTCTGTGATGCCCTTAGATGTTCTGGCCCGCACGCCGCTACACTGAAGGGATCAACGTGTTCT  
CCCCCTCCGAGAGGAGCGGGTAACCCGATCAAAAACCTTCATGATAGGGATTGGGGCTTGCAATTTGTTCCCATGAACGAGGAATT  
CCCAGTAAGCGCAAGTCAATCAGCTTGGCTTGGATTACGTCCCTGCCCTTTGTACACACCGCCCGTCCCTACTACCGATTGAATGATT  
TAGTAGGCCTTCGGACTGGCGCTCTTGTTGTTCTACCCCATCCCTTCTATTCTCGCAAGGGTCTGGAGGTTGGGTTCTCGCC  
TCGAGCTGACGGAAAGATGTCCAAACTTGATCATTTAGAGGAAGGAAAGTTCGTAACAAGGTTTCCGTAGGTGAACCTGC

d) 18s rRNA sequence of *Panulirus homarus* from Chinnamuttom

ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCAATGGATCTGTAAACCCACTTACTTGGATAACTGTGGTAATT  
CTAGAGCTAATACATGCATCACGTCTCTGACCGCAAGGGAAGAGCGCTTTTATTAGTTCAAAACCCAGTCGGGCCTCGGTCCGTAC  
CCACCTGTGGTGAATCTGAATAACTTCTCGCTGAGCGCAGGGTCTACGCACCGGCCCGAGTCTTTCAGTGTCTGCCTTATCAGC  
TTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTAACGGGGAATCAGGGTTCGATTCCGGAGAGGGAGCCTGAGAAACG  
GCTACCACATCTAAGGAAGGCAGCAGGCACGCAAATTAACCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAACGATGCGAGAC  
TCATCCGAGGCCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGAGGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCG  
CGGTAATCCAGCTCCAATAGCGTATATAAAGTTGTTGCGGTTAAAAAGCTCGTAGTTGGATTTTCAGTCCCGACTGACGGTTCA  
CCGCCCGGTGTCTACTGTACGCTCCGAACAGCCGCACCGCCGGCTCGCACGGGGTGTCTTTCATCGAGTGTCCCGAGTGGCCGGC  
TCGTTTACTTTGAAAAAATTAGAGTGTCTCAGAGCAGGCTCCTTGAATGGCCTGAATGTCTATGCATGGAATAATGGAATAGGACCT  
CGGTTCTATTTTGTGGTTTACGGAACCCGAGGTAATGACTAATAGGAACAGGCGGGGTCATTTCGATTTGCGACGCTAGAGGTGAA  
ATTCTTGGACCGTTCGCAAGACAACACTGCGAAAGCAATTTGCCAAGGATGTTTTCATTAATCAAGAACGAAAGTTAGAGGTTCGAAG  
GCGATCAGATACCGCCCTAGTTCTAACCATAAACGATGCTGACTAGCGATCCGCCGGCGTATTCCCATGACCCGGCGGGCAGCTT  
CCGGAAACCAAGTCTTTGAGTTCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAAGGGCACCACAGGA  
GTGGAGCCTGCGGCTTAAATTTGACTCAACAGGGGAACCTCACCAGGCCCGGACACCGGAAGGATTGACAGATTGAGAGCTCTTTCT  
CGATTCCGGTGGGTGGTGGTGCATGGCCGTTCTTAGTTGGTGGAGCGATTGTCTGGTTAATTCGGATAACGAACGAGACTCTAGCC  
TATTAAGTGTGACGGATCTCCAGCATGCTCCGTGGTGTCCGTTGCAACGCTCTTCTTAGAGGGATAAGCGGCAATCTAGCCGC  
ACGAAATGAGCAATAACAGGTCTGTGATGCCCTTAGATGTTCTGGCCCGCACGCCGCTACACTGAAGGGATCAACGTGTTCTCC  
CCCTCCGAGAGGAGCGGGTAACCCGATCAAAAACCTTCATGATAGGGATTGGGGCTTGCAATTTGTTCCCATGAACGAGGAATTCC  
CAGTAAGCGCAAGTCAATCAGCTTGGCTTGGATTACGTCCCTGCCCTTTGTACACACCGCCCGTCCCTACTACCGATTGAATGATTTA  
GTGAGGCCTTCGGACTGGCGCTCTTGATGTTCTACCCCATCCCTTCTATTCTCGCAAGGGTCTGGAGGTTGGGGTTCTCGCCCT  
GAGCTGACGGAAAGATGTCCAAACTTGATCATTTAGAGGAAGTAGAGTTCGTAACAAGGTTTCCGTAGGTGAACCTGC

e) 18s rRNA sequence of *Palinurus homarus* from Tuticorin

ATTAAGGC GAAACCGCGAATGGCTCAT TAAATCAGCTATGATT CATTGGATCTGTAAACCCACTTACTTGGATAACTGTGGTAATT  
CTAGAGCTAATACATGCATCACGTCTCTGACCGCAAGGGAAGAGCGCTTTTATTAGTTCAA AACCCAGTCGGGCCTCGGTCCGTAC  
CCACCTGTGGTGAATCTGAATAACTTCTCGCTGAGCGCAGTGTCTACGCACCGGCCGAGTCTTTCAAGTGTCTGCCTTATCAGC  
TTTCGATTGTAGTTATGCGCCTACAATGGCTATAACGGGTAACGGGGAATCAGGGTTCGATTCCGGAGAGGGAGCCTGAGAAACG  
GCTACCACATCTAAGGAAGGCAGCAGGCACGCAAATTACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAACGATGCGAGAC  
TCATCCGAGGCCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGAGGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCG  
CGGTAATTCAGCTCCAA TAGCGTATAT TAAAGTTGTTGCGGTTAAAAAGCTCGTAGTTGGATTTCAGTTC CGGACTGACGGTTCA  
CCGCCCGGTGTCTACTGTCACGCTCCGAACAGCCGCACGGCCGGCTCGCACGGGGTGTCTTTCATCGAGTGTCCCGAGTGGCCGGC  
ACGTTTACTTTGAAAAAATTAGAGTGTCTCAGAGCAGGCTCCTTGAATGGCCTGAATGTCTATGCATGGAATAATGGAATAGGACCT  
CGGTTCTATTTTGGTTGGGTTGT CGGAACCCGAGGTAATGACTAATAGAAACAGGCCGGGGCATTCTGATTCGACGCTAGAGGTGA  
AATTC TTGACCGTCGCAAGACGAAC TACTGCGAAAGCATTTGCCAAGGATGTTTCA TTAATCAAGAACGAAAGTTAGAGGTT CGA  
AGGGATCAGATACCGCCCTAGTTCTAACCATAAACGATGCTGACTAGCGATCCGCCGGCGTTATTC CCATGACCCGGCGGGCAGC  
TTCGGGAAACCAAAGTCTTTGAGTTCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAAGGGCACCCCCAG  
GAGTGGAGCCTGCGGC TTAATTTGACTCAAAACGGGGAACCTCACCAGGCCCGACACCGGAAGGATTGACAGATTGAGAGCTCTT  
TCTCGATTCCGGTGGGTGGTGGTGCATGGCCGTTCTTAGTTGGTGGAGCGATTTGTCTGGTTAATTC CGATAACGAACGAGACTCTA  
GCCTATTA ACTAGTCGACGGATCTCCAGCATGCTCCGTGGTGTCCGTTGCGAACGTC TTTCTAGAGGGATAAGCGGCAATTTCTAGC  
CGCACGAGAATGAGCAATAACAGGTC TGTGATGCCCTTAGATGTTCTGGGCCGCACGCGCTACACTGAAGGGATCAACGTGTTT  
TCCCCCTCCGAGAGGAGCGGGTAACCCGATCAAAACCC TTTATGATAGGGATTGGGGCTTGCAATTGTTTCCCATGAACGAGGAAT  
TCCCAGTAAGCGCAAGTCATCAGCTTGC GTTGATTACGTCCCTGCCCTTTGTACACACCGCCCGTCTACTACCGATTGAATGAT  
TTAGTGAGCCTTCGGACTGGCGCTCTTGAATGTTCTACCCCATCCCTTCTATTCTCGCAAGGGTCTTGAGGTTGGGGTTCTCGC  
CTCGAGCTGACGGAAGATGTCCA AACTTGATCATTTTAGAGGAAGTAAAGTCGTAACAAGGTTTCCGTAGGTGAACCTGC

Plate: 2.

ClustalW 2.0 Multiple Sequence alignment and Phylogenetic distance tree using UPGMA method

Muttom ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCATTGGATCTGTAAACC  
Chinnamuttom ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCATTGGATCTGTAAACC  
Cochin ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCATTGGATCTGTAAACC  
Tuticorin ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCATTGGATCTGTAAACC  
Vizhijam ATTAAGGCGAAACCGCGAATGGCTCATTAAATCAGCTATGATTCATTGGATCTGTAAACC  
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Muttom CACTTACTTGGATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Chinnamuttom CACTTACTTGGATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Cochin CACTTACTTGGATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Tuticorin CACTTACTTGGATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Vizhijam CACTTACTTGGATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
\*\*\*\*\*

Muttom AAGGGAAGAGCGCTTTTATTAGTTCAAAACCAAGTCGGGCCTCGGTCCGTCAACCCCTGT  
Chinnamuttom AAGGGAAGAGCGCTTTTATTAGTTCAAAACCAAGTCGGGCCTCGGTCCGTCAACCCCTGT  
Cochin AAGGGAAGAGCGCTTTTATTAGTTCAAAACCAAGTCGGGCCTCGGTCCGTCAACCCCTGT  
Tuticorin AAGGGAAGAGCGCTTTTATTAGTTCAAAACCAAGTCGGGCCTCGGTCCGTCAACCCCTGT  
Vizhijam AAGGGAAGAGCGCTTTTATTAGTTCAAAACCAAGTCGGGCCTCGGTCCGTCAACCCCTGT  
\*\*\*\*\*

Muttom GGTGAATCTGAATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Chinnamuttom GGTGAATCTGAATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Cochin GGTGAATCTGAATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Tuticorin GGTGAATCTGAATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
Vizhijam GGTGAATCTGAATAAAGTGTGGTAATTCTAGAGCTAATACATGCATCACGTCCTGACCGC  
\*\*\*\*\* \*\* \*\* \*\*\*\*\*

Muttom AGTGTCTGCCTTATCAGCTTTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTA  
Chinnamuttom AGTGTCTGCCTTATCAGCTTTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTA  
Cochin AGTGTCTGCCTTATCAGCTTTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTA  
Tuticorin AGTGTCTGCCTTATCAGCTTTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTA  
Vizhijam AGTGTCTGCCTTATCAGCTTTTCGATTGTAGGTTATGCGCCTACAATGGCTATAACGGGTA  
\*\*\*\*\*

Muttom ACGGGGAATCAGGGTTTCGATTCGCGGAGAGGGAGCCTGAGAAACGGCTACCATCTAAGG  
Chinnamuttom ACGGGGAATCAGGGTTTCGATTCGCGGAGAGGGAGCCTGAGAAACGGCTACCATCTAAGG  
Cochin ACGGGGAATCAGGGTTTCGATTCGCGGAGAGGGAGCCTGAGAAACGGCTACCATCTAAGG  
Tuticorin ACGGGGAATCAGGGTTTCGATTCGCGGAGAGGGAGCCTGAGAAACGGCTACCATCTAAGG  
Vizhijam ACGGGGAATCAGGGTTTCGATTCGCGGAGAGGGAGCCTGAGAAACGGCTACCATCTAAGG  
\*\*\*\*\*

Muttom AAGGCAGCAGGCACGCAAAATACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAAC  
Chinnamuttom AAGGCAGCAGGCACGCAAAATACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAAC  
Cochin AAGGCAGCAGGCACGCAAAATACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAAC  
Tuticorin AAGGCAGCAGGCACGCAAAATACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAAC  
Vizhijam GAGGCAGCAGGCACGCAAAATACCCACTCCCGGCACGGGGAGGTAGTGACGAAAAATAAC  
\*\*\*\*\*

Muttom GATGCGGAGACTCATCCGAGGCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGA  
Chinnamuttom GATGCGGAGACTCATCCGAGGCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGA  
Cochin GATGCGGAGACTCATCCGAGGCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGA  
Tuticorin GATGCGGAGACTCATCCGAGGCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGA  
Vizhijam GATGCGGAGACTCATCCGAGGCTCGCAATCGGAATGAGTACACTTTAAATCCTTTAACGA  
\*\*\*\*\*

Muttom GGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAAATCCAGCTCCAATAGCGT  
Chinnamuttom GGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAAATCCAGCTCCAATAGCGT  
Cochin GGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAAATCCAGCTCCAATAGCGT  
Tuticorin GGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAAATCCAGCTCCAATAGCGT  
Vizhijam GGATCCATTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAAATCCAGCTCCAATAGCGT  
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Tuticorin ATATTAAGTTGTTGCGGTTAAAAAGCTCGTAGTTGGATTCAGTTCGGACTGACGGTT  
Vizhijam ATATTAAGTTGTTGCGGTTAAAGCTCGTAGTTGGATTCAGTTCGGACTGACGGTT  
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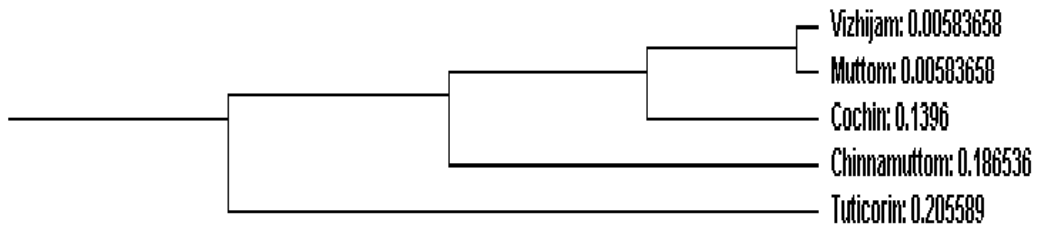
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Cochin TACCGATTGAATGATTTAGTGAGGCCTTCGGACTGGCGCTCTTGGATGTCTACCCCATC  
Tuticorin TACCGATTGAATGATTTAGTGAGGCCTTCGGACTGGCGCTCTTGAATGTCTACCCCATC  
Vizhijam TACCGATTGAATGATTTAGTGAGGCCTTCGGACTGGCGCTCTTGGATGTCTACCCCATC  
\*\*\*\*\*

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Tuticorin CCTTCTATTCTCGCAAGGGTCTGGAGGTTGGGGTTCTCGCCTCGAGCTGACGGAAAGAT  
Vizhijam CCTTCTATTCTCGCAAGGGTCTGGAGGTTGGGGTTCTCGCCTCGAGCTGACGGAAAGAT  
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Vizhijam GTCCAAACTTGATCATTAGAGGAAGTAAAGTCGTAACAAGGTTTCCGTAGGTGAACCTG  
\*\*\*\*\*

Muttom C  
Chinnamuttom C  
Cochin C  
Tuticorin C  
Vizhijam C  
\*

**Fig: 12**  
**Phylogenetic tree and genetic distance based on 18S rRNA Sequence.**





## Discussion

Characters of DNA sequences have been known to affect topological resolution in phylogenetic analysis. The most common such character is site saturation by multiple substitutions that may cause obscurity in true relationships of taxa in evolution (Swofford *et al.*, 1996; Jesus *et al.*, 2007). However, there is the lack of saturation of transitions for the 16S of the freshwater prawns (Murphy and Austin, 2005; Liu *et al.*, 2007). In our study, there was deceleration in transitions in 18S rRNA sequences. Increasing a sequence length increases the number of phylogenetic informative sites that improves the resolution of phylogenetic analysis (Miyamoto *et al.*, 1990; Chang *et al.*, 2005). It is correctly said that increasing a sequence length provides more data in the analysis, the better the chance to obtain a better phylogeny. Liu *et al.*, (2007) combined the 16S sequences and the COI sequences to make the combined sequences 37% larger in the length and 47% more in the total parsimonious information sites than those of the 28S. The combined sequences improved the resolution of the phylogenetic tree derived from the 16S, but many of the internal multifurcating nodes were unresolved and the phylogenetic relationships of many taxa remained obscure. Obviously, transitional saturation and sequence length were not the primary factors for the 16S to produce poor resolution in the phylogenetic analysis for the freshwater prawns.

Murphy and Austin (2005) attributed this poor resolution in phylogenetic analysis of freshwater prawns from the 16S to a rapid and explosive radiation in their early evolutionary history, which might have occurred during the Oligocene or early Miocene. They speculated

that was a period with the proper environments for rapid dispersion and diversification of amphidromous prawns in the Southeast Asia and farther to Australia, the western Pacific islands, India, Africa, and even Atlantic coasts of Tropical America (Chen *et al.*, 2009). The rapid radiation created numerous taxa within a short time period. The 18S exhibits small genetic distance range, resulting in a severe convergence of the taxa within the range so that any bifurcating branching patterns could not be determined unambiguously, as expressed by the presence of severe polytomies in the phylogenetic trees.

Phylogenetic information in DNA sequences is built up by nucleotide substitutions over time. Due to the limited number of character states, phylogenetic signals also erode by similar process. Several evolutionary factors can potentially mislead phylogeny estimations (Conant and Lewis, 2001), such as heterogeneity in substitution rates among lineages (Felsenstein, 1978) and sites (Navidi *et al.*, 1991; Sidow and Steel, 1992; Yang, 1993), transition/transversion bias (Kimura, 1980; Wakeley, 1993), nonindependence of sites within a gene (Goldman and Yang, 1994; Schoniger and Von Haeseler, 1995; Muse, 1995, 1996), and non-stationarity of nucleotide frequencies across lineages (Loomis and Smith, 1990; Burggraf *et al.*, 1992; Hasegawa and Hashimoto, 1993; Lockhart *et al.*, 1994; Galtier and Gouy, 1995, 1998).

The gamma distribution has been proposed to model variable rates at sites (Jin and Nei, 1990; Li *et al.*, 1990; Tamura and Nei, 1993). The distribution involves a shape parameter of which the value is inversely related to the extent of rate variation at sites. Sullivan *et al.*,(1995) and Misof *et al.*,(2001) demonstrate that there is extreme among-site variation in mitochondrial 12S sequences, which has a highly skewed distribution of the rates

and makes the sequences particularly susceptible to the misleading effects of non-independency and other nonrandom noise in phylogenetic analysis.

The 18S had a-value less than 0.5, the presence of extremely high among-site rate variation (Tateno *et al.*, 1994; Sullivan *et al.*, 1995), whereas the 28S had the value higher than 0.5, indicating very low among-site rate variation. The lower a-value indicated higher among-site variation rates (Page and Holmes, 1998). The higher among site rate variation of the 16S as indicated by its low value suggested that most sites of its sequences were invariable, but a few had very high substitution rates. Consequently, it was less informative, and thus the divergences among sequences were smaller (Yang, 1996).

On the basis of Phylogenic tree highest genetic distance was found in Tuticorin population (0.205589) and less genetic distance found in Vizhinjam population (0.00583658). Phylogenic tree produced mainly two clusters. First cluster contains Tuticorin populations and the second cluster contains remain four populations. The results of multiple sequence alignment of 18S gene sequence in five different collection spots were showed that the base pairs varied in all populations.

The combined analysis of different DNA segments has been often tested, but the results are controversial (Brower *et al.*, 1996; Wiens, 1998; Lecointre and Deleporte, 2005; Hasegawa and Kasuya, 2006). A combined analysis is effective if the partitioned components of the data sets are congruent with consistent phylogenetic signals (Chavarria and Carpenter, 1995; Hasagawa and Kasuya, 2006). In this study, the partition-homogeneity

test showed that the 18S data sets were congruent; nevertheless, the combined analysis with ML, MP and MENJ showed no improvement from the 28S, but with BI produced mixed results. Obviously, in this instance, the combined analysis is not the first choice in this phylogenetic analysis; the separate analyses should be done first (Lecointre and Deleporte, 2005; Hasegawa and Kasuya, 2006). To find the most plausible phylogenetic hypothesis, a mutual reference should be made among the results of the separate and combined analyses (Hasegawa and Kasuya, 2006).