

## CHAPTER 4

### MATERIALS AND METHODS

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#### 4.1 MATERIALS

Potassium ferrocyanide ( $K_4 [Fe(CN)_6]$ ), Potassium ferricyanide ( $K_3[Fe(CN)_6]$ ), Sodium bicarbonate ( $NaHCO_3$ ) and Potassium Chloride (KCl) used were of analytical grade and were purchased from Merck, Germany. 99 % of L- $\alpha$ -Phosphatidylcholine was purchased from Sigma Aldrich. Stock solution of L- $\alpha$ -Phosphatidylcholine (50 mg/mL) was prepared by using Analytical Grade chloroform. BLM forming dispersions were prepared in n-Decane (Merck, Germany). All the aqueous solutions used in our studies were prepared using Milli-Q water ( $R > 18 M\Omega \text{ cm}^{-1}$ ).

#### 4.2 DRUG FOR THE STUDIES - LORAZEPAM

Lorazepam was extracted from commercially available calmese tablets, using 90% ethanol solution, in its hydrochloride form. It is precipitated by adding excess water to the 90% ethanolic solution. The precipitate was separated by filtration, dried and used for studies.

#### 4.3 CONSTRUCTION OF BLM CHAMBERS

Interaction of Lorazepam with planar lipid membranes was studied using indigenously constructed BLM chambers. Two 24 mm diameter cavities, each of 5ml capacity were drilled in a polished PMMA block of dimension 85 x 60 x 22mm. The cavities are exactly aligned and almost touch

each other. The cavities were drilled in a computerized numerical control lathe. At the meeting point of the cavities the block was cut into two halves. A hole of diameter 1.34 mm was drilled in a 1mm thick PMMA sheet and chamfered using a 1.25 mm drill bit from both sides to equal depths to form the membrane-supporting aperture. The aperture was examined through a travelling microscope with a magnification of 10 x and its diameter was measured. The septum having perfect circular aperture with minimum boundary ripples was used for the studies. This septum was inserted between the two halves of the chamber blocks such that the aperture is at one-third distance from the bottom of the cavity and glued with utmost care. The photograph of BLM chamber is shown in the following Figure.4.1.



**Figure 4.1** The BLM chamber

#### **4.3.1 Vibration Isolation Platform**

The interferences from the floor vibrations are arrested by using a vibration isolated platform (MINUSK USA).

#### **4.3.2 Fabrication of Electrodes**

A two-electrode setup was used for studying the interaction of Lorazepam with planar or black lipid membrane. The Silver-Silver Chloride electrodes were fabricated by following standard procedures. Silver wire of 1mm diameter was anodized in 0.1M KCl solution to form the AgCl coating over the wire (Geddes, 1972) (Corvington, 1979). In AC measurements the two electrodes used are Ag/AgCl electrodes. The electrical connectivity between the electrolytic solution in the cavities and electrode immersed solutions was given by U shaped salt bridges made of glass filled with 1.5% agar-agar gel and saturated with KCl.

A three electrode assembly was used for studying interaction of Lorazepam with sb-BLM ( salt bridge supported BLM) in KCl bath solutions where in a Pt foil served as the counter electrode, a Ag wire coated with AgCl served as the reference electrode, while the BLM formed on the agarose gel surface served as the working electrode. 0.3 g of agarose (Sigma Aldrich) was dissolved in 15 mL of boiling saturated KCl solution and the Teflon tubes were immersed so that the agar gel filled the tubes completely. Ag-AgCl reference electrode was prepared by anodizing Ag wire dipped in 0.1 M KCl solution. The anodized Ag - AgCl electrode was inserted into the agar gel

filled Teflon tube. The end of the Teflon tube was cut using a very sharp knife and 3  $\mu\text{L}$  of lipid solution in n-decane was applied on the fresh surface of the agarose gel and allowed to dry under nitrogen atmosphere.

Studies on glassy carbon supported BLM, in the presence of marker ions were also made using a three electrode set up. The glassy carbon electrode of diameter 2 mm was used as a solid support for bilayer lipid membrane. The surface of glassy carbon electrode was polished with alumina slurry using polishing kit, washed with double distilled water and activated electrochemically by cyclic voltammetric method using the procedure described by Balamurugan et al (Balamurugan et al, 2013) and used for studies. All electrochemical experiments, using GCE-BLM, were performed with a potentiostat, GAMRY REFERENCE 3000. BLM covered GCE served as working electrode, Ag/AgCl electrode served as reference electrode and a Pt foil was used as the counter electrode.

### **4.3.3 Faraday Cage**

The currents involved in the BLM experiments are extremely low, which are of the order  $10^{-12}$  A. Any stray current pick-up will greatly affect the readings. Hence, the BLM set-up (the chamber electrode, stirring units etc) was placed in a metal cage called Faraday cage (Hanke and Schlue, 1993) (Sakmann and Neher, 1983) and the cage was grounded. At most care was taken to avoid stray-current leakage between the two aqueous chambers due to cracks in the septum in the case of black lipid membrane studies.

#### **4.4 PREPARATION OF BLM FORMING DISPERSION**

100 $\mu$ L of chloroform stock solution of the L- $\alpha$ -Phosphatidylcholine (50 mg/mL) was taken in a screw - cap bottle and nitrogen gas was purged forcibly into the tube in order to form a dry thin film of phospholipid and to evaporate chloroform. The phospholipid film was then immediately dispersed in 200 $\mu$ l of n-decane and used as the BLM forming dispersion.

#### **4.5 FORMATION OF MODEL BLM SYSTEMS**

##### **4.5.1 Formation of planar or black lipid membrane**

About 1 $\mu$ L of the lipid dispersion in n-decane was applied on the septum aperture as a preconditioning step (Tien, 1974) and dried. Then required bathing solutions were taken in both compartments of the BLM chamber. Salt bridges were placed on both the sides. After placing the block on a stand electrical connections were made. Approximately 5 $\mu$ L of the BLM forming lipid dispersion was taken in a microlitre syringe. The tip of the syringe was immersed in the bath solution and held touching the aperture. The lipid dispersion was then delivered directly over the aperture. The dispersion spreads and sticks to the aperture as a lens and then slowly thins down. The solvent molecules were slowly excluded and the phospholipid molecules self assemble into a bilayer to form planar or black lipid membrane.

Brushes were used by workers previously to apply lipid dispersions over the aperture and this technique was found to associate with lot of uncertainties. To avoid the troubles caused by such uncertainties, H.Ti.Tien suggested a more

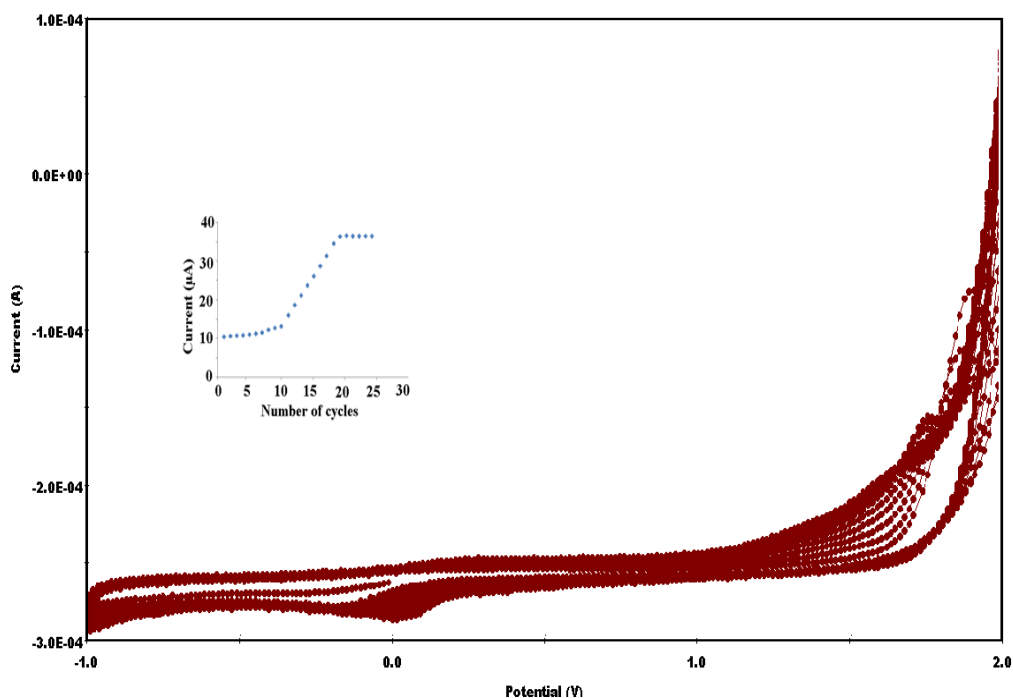
elegant technique (Tien, 1974) where syringe is used to deliver the lipid dispersion at the aperture which is called syringe technique. Besides this a number of techniques are available for the formation of BLMs (Hanke and Schlue, 1993) (Tien, 1974). Being simple and suitable to form BLM at the aperture between two cavities; the syringe technique was used throughout the work.

#### **4.5.2 Formation of sb-BLM**

The end of the Teflon tube filled with agar gel saturated with KCl was cut by a very sharp knife and 3  $\mu\text{L}$  of the lipid dispersion in n-decane was applied on the fresh surface and allowed to dry under nitrogen atmosphere. Again 3  $\mu\text{L}$  of the lipid solution was applied and immersed in 3.5 mL NaCl bath solution for 30 minutes, where the lipid (L- $\alpha$ -Phosphatidylcholine) molecules self assemble into a lipid bilayer.

#### **4.5.3 Formation of solid supported BLM (GCE-BLM)**

Using cyclic voltammetric technique the surface of GCE was activated in 0.1 M  $\text{NaHCO}_3$  solution. Placing Ag/AgCl electrode as reference electrode, a Pt foil as counter electrode, the surface of glassy carbon electrode was activated keeping it as working electrode in 0.1 M  $\text{NaHCO}_3$  solution by cyclic voltammetry. The glassy carbon electrode was mechanically polished well before electrochemical activation using alumina slurry. The cyclic voltammograms were recorded in the potential range -1.0 to 2.0 V at the scan rate of 100 mV/s. The cyclic voltammograms recorded during electrochemical activation of GCE is shown in Figure 4.2.



**Figure 4.2** Cyclic voltammograms obtained during electrochemical activation of GCE

The formation of highly porous, highly reactive oxygen containing functional groups and oxidized carbon on the GCE surface is related to the anodic peak observed around 1.8 V (Ilangovan and Chandrasekar pillai, 1999). This anodic peak current increased significantly after 10 cycles and reached a maximum at 21<sup>st</sup> cycle (Inset of Figure 4.2). Hence, before forming BLM on GCE, it was electrochemically activated atleast by 20 voltammetric cycles in the above mentioned potential range.

The electrochemical activation of GCE provided highly porous, rough and highly electrochemically active surface compared to that of bare GCE. 3 μL of lipid (L- $\alpha$ -Phosphatidylcholine) dispersion in n-decane was applied on the dry electrochemically activated GCE surface using a micro syringe and



immediately immersed in the KCl bath solution, where the phospholipid molecules undergo self assembly spontaneously into a bilayer lipid membrane. The electrical properties of s-BLMs attained stable values after 30 minutes. This indicates the formation of a stable membrane on the electrochemically activated GCE surface.

The interaction between Lorazepam and s-BLM in the presence of marker (ferri-ferro cyanide) ions was studied in KCl bath solutions at pH 7, using BR buffer. pH 7 BR buffer was prepared by adding 57.2 mL of 0.2 N NaOH to 100 mL of pH 1.86 BR buffer. BR buffer solution of pH 1.86 was prepared by adding 2.4761 g of boric acid to a 1.0 L solution containing 2.3 mL of 17.4 N acetic acid and 2.7 mL of 14.7 N phosphoric acid.

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