MATERIALS AND METHODS

Meliolaceous fungi are predominantly foliicolous and rarely infect the soft stems and tender shoots. Collections of these fungi are easier than that of the fleshy fungi. While collecting the infected plant parts, field notes were made regarding their pathogenicity, nature of colonies, nature of infection, locality, altitude, etc. For each collection separate field number was given. In the field, such infected plants were collected separately in polythene bags along with the host twig (preferably with the reproductive parts to facilitate the corresponding host identity). These infected plant parts were pressed neatly and dried in-between blotting papers. After ensuring their dryness they were kept in the manifold or butter paper folders. Later these folders were placed in thick paper envelopes of convenient size with the name of the host, locality, date of collection, place of collection, name of the collector along with the field number written on the top corner. These envelopes were serially arranged in a rack based on their collection number. Friction between the envelope and the material was avoided to keep the mycelia, perithecia and setae intact. Such materials were used for the microscopic study.

For microscopic study, scrapes were taken directly from the infected host and mounted in 10% KOH solution. After 30 minutes, KOH was replaced by Lacto phenol (prepared according to Rangaswamy, 1975). Both the mountants worked well as clearing agents and made the septa visible for taking measurements.

To study the entire colony in its natural condition, a drop of high quality natural coloured or well transparent nail polish was applied to the selected colonies and carefully thinned with the help of a fine brush without disturbing the colonies. Colonies with hyperparasites showed wooly nature and were avoided. The treated colonies along with their host plants were kept in dust free chamber for half an hour. When the nail polish on the colonies dried fully, a thin, colourless or slightly apple rose coloured (depending up on the colour tint of the nail polish) film or flip was formed with the colonies firmly embedded in it. In case of soft host parts lifts the flip up with a slight pressure on the opposite side of the leaves and just below the
colonies. In case of hard host parts the flip was eased off with the help of a razor or scalpel. A drop of DPX was put on a clear slide and the flip was spread properly on it. One or two more drops of DPX was again added on the flip and a clean cover glass was placed over it and a gentle pressure on the cover glass brings out the excess DPX and it was removed after drying. Care was taken to avoid air bubbles. These slides were labelled and placed in a dust free chamber for 1-2 days for drying. These permanent slides were then used for further studies.

Nikon Alphaphot - 2 microscope was used for studying and microphotographs of materials. Mirror type Camera Lucida used to make line drawings. Earlier materials collected from these area and described in TBGT (Tropical Botanic Garden and Travancore Herbarium) were studied and recent materials were also deposited in TBGT and HCIO (Herbarium Cryptogamae Indiae Orientalis) after the study.