Standard curve of Na-pyruvate
(Enzymes: ALT, AST)

N = 3

OD 505 nm

Conc. (ug)

Standard curve of p-aminophenol
[Enzyme: Aniline hydroxylase]

N = 3

OD 630 nm

Conc. (ug)
Standard curve of GSH
[Reduced GSH, Enzyme: GSH-Px]

\[ N = 3 \]

\[
\begin{array}{c|c}
\text{Concn (ug)} & \text{OD 412 nm} \\
\hline
0 & 0.00 \\
5 & 0.10 \\
10 & 0.20 \\
15 & 0.30 \\
20 & 0.40 \\
25 & 0.50 \\
30 & \\
\end{array}
\]

Standard curve of p-nitroaniline
[Enzyme: Gamma-GT]

\[ N = 3 \]

\[
\begin{array}{c|c}
\text{Conc. (ug)} & \text{OD 405 nm} \\
\hline
0 & 0.00 \\
2 & 0.20 \\
4 & 0.40 \\
6 & 0.60 \\
8 & 0.80 \\
10 & \\
\end{array}
\]
Standard curve of MDA
(Lipid Peroxidation)

N = 3

OD 535 nm

Conc. (nmoles)

Standard curve of p-nitrophenol
[Enzyme: ALP]

N = 3

OD400

Concn. (ug)
Preparation of collagen from rat tails

Collagen was isolated from rat tails according to the modification of Gebhart and Jung et al., (1982). Cut tails, preserved at -80 °C were cleansed with cotton swab soaked in 70% ethanol and cut into pieces with a plier. Tendons were pulled out, air dried and exposed to UV radiation for one hour. Dry tendons (600 mg) were soaked in 200 ml of 10% NaCl, w/v, for 2-3 days. The solution was changed each day. Thereafter, the tendons were soaked in 200 ml, 70 mM K₂HPO₄ for 2-3 days, changing the solution each day. Subsequently, the tendons were kept under diethylether (100ml) for one day, and finally suspended in 100 ml (1%, v/v) acetic acid at room temperature, under constant stirring for 2-3 days. The collagen solution was filtered through muslin cloth and dialysed against 2 litres of 70 mM potassium phosphate buffer, pH 7.6 at 4°C, till collagen precipitated in the dialysis bags. The precipitate was dissolved in 1% acetic acid and diluted 1:4 with 0.1 % acetic acid. The collagen solution was exposed to γ-radiation, 4000 rads at 4°C and stored at -80 °C.